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REVIEW

Exercise as a tool to mitigate metabolic disease

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Abstract

Metabolic diseases, notably obesity and type 2 diabetes (T2D), have reached alarming proportions and constitute a significant global health challenge, emphasizing the urgent need for effective preventive and therapeutic strategies. In contrast, exercise training emerges as a potent intervention, exerting numerous positive effects on metabolic health through adaptations to the metabolic tissues. Here, we reviewed the major features of our current understanding with respect to the intricate interplay between metabolic diseases and key metabolic tissues, including adipose tissue, skeletal muscle, and liver, describing some of the main underlying mechanisms driving pathogenesis, as well as the role of exercise to combat and treat obesity and metabolic disease.

adipose tissue; insulin resistance; liver; metabolism; skeletal muscle

INTRODUCTION

The growing prevalence of metabolic diseases, including obesity, type 2 diabetes (T2D), and hypertension, among others, has reached alarming proportions in recent decades, becoming a global burden (1). Specifically, regarding obesity and type 2 diabetes, it is estimated that more than 650 million adults worldwide have obesity (body mass index $\geq 30~{\rm kg/m^2}$), and over 537 million adults are living with type 2 diabetes (2, 3). Substantial evidence has shown that sedentary behavior or insufficient levels of physical activity are key factors involved in the development of metabolic diseases and contribute to shortened life expectancy (4–6).

Exercise training has a central role in the prevention and treatment of chronic diseases, including obesity and T2D. There are numerous beneficial effects and adaptations of exercise for metabolic health including, but not limited to, improvements in glucose tolerance, insulin sensitivity, redox health, adaptations to the gut microbiota, and reduced inflammation (7–10). Furthermore, positive adaptations to exercise are observed in several metabolic tissues, notably in skeletal muscle, adipose tissue, and liver (11). In this review, we will discuss the relationship between obesity and type 2 diabetes, how these conditions affect metabolic tissues, and how exercise-induced adaptations in the adipose tissue, skeletal muscle, and liver improve metabolic health.

THE RELATIONSHIP BETWEEN OBESITY AND TYPE 2 DIABETES AND THE EFFECTS OF EXERCISE

It is well established that there is a significant relationship between obesity and the development of insulin resistance in peripheral tissues and, consequently, type 2 diabetes. There are several proposed mechanisms involved in this process, including inflammation, increased levels of free fatty acids in the circulation, and mitochondrial dysfunction, all of which may play an important role.

In contrast, there are numerous adaptations in response to exercise that can combat obesity and metabolic disease. This includes improvements in blood pressure, circulating lipid profile, inflammatory profile, cardiorespiratory fitness, and cardiovascular biomarkers, among others (12–15). Specifically regarding obesity, exercise training is considered an effective strategy for weight and adiposity management, and it is associated with a reduction in body mass, adiposity, and cardiometabolic risk factors (16–18). In addition, regular physical activity has a profound impact on diabetes management and it is associated with enhanced insulin sensitivity, pancreatic β -cell function, and improved whole body glucose metabolism (13, 19–21), which directly promotes the improvement of glycemic control (7, 22).

The positive benefits observed with regular physical activity and exercise, especially those related to improvement in metabolic health aspects, occur primarily through adaptations to the adipose tissue, skeletal muscle, and liver. Therefore, in the next sections, we will describe the main mechanisms linking obesity, insulin resistance, and type 2 diabetes in peripheral tissues, and define some of the main exercise-induced adaptations to these metabolic tissues with a focus on their preventive and therapeutic role in metabolic health.

ADIPOSE TISSUE

Adipose tissue is a highly dynamic tissue and has important functions, including energy storage in the form of







triglycerides, protection against mechanical stress, the release of hormones and energetic substrates, among others (23). Adipose tissue can be broadly classified into two different types: white adipose tissue (WAT), whose primary functions are energy storage and insulation, and can be subdivided into subcutaneous WAT (scWAT) and visceral WAT (vWAT); and brown adipose tissue (BAT), which is a metabolically active tissue involved in thermogenesis by uniquely expressing uncoupling protein 1 (UCP1) (24–27).

Adipose Tissue, Obesity, and Inflammation

Obesity is a chronic and complex disease characterized by an abnormal or excessive accumulation of adipose tissue. The increase in WAT mass, especially vWAT, is closely associated with the development of insulin resistance in metabolically hormone-responsive tissues, such as skeletal muscle, liver, and adipose tissue itself (28, 29). In addition, an excessive amount of WAT is associated with the release of free fatty acids, glycerol, several proinflammatory cytokines and chemokines, hormones, and other factors that are closely involved in the development of insulin resistance (30, 31).

The mechanisms linking obesity to insulin resistance and predisposition to T2D are numerous. One proposed mechanism is obesity-related inflammation. Adipose tissue contains multiple immune cells that together maintain the integrity and hormonal sensitivity of adipocytes (32). A class of immune cells present in adipose tissue are macrophages, which are critical contributors to inflammation and insulin resistance. In obesity, the number of adipose tissue macrophages increase and comprise up to 40% of all adipose tissue cells, which can be seen histologically by the formation of crown-like structures, consisting of macrophages surrounding dead adipocytes (33). Adipose tissue macrophages (ATMs) can be either pro- or anti-inflammatory and are typically termed as M1-like or as M2-like, respectively (34). The terms M1-like and M2-like are used to generally depict the proinflammatory state of recruited ATMs versus the antiinflammatory state of resident ATMs (35). The M2-like ATMs secrete anti-inflammatory cytokines such as interleukin-10 (IL-10) and contribute to the maintenance of insulin sensitivity and adipose homeostasis (36, 37). Conversely, M1-like ATMs secrete proinflammatory cytokines including tumor necrosis factor-alpha (TNF- α), IL-1 β , and IL-6 (36). In addition, the adipose tissue itself also secretes several proinflammatory adipokines/cytokines including TNF-α, IL-6, among many others, which leads to activation of classical inflammatory signaling such as c-Jun amino-terminal kinase (JNK) and nuclear factor-κB (NF-κB) pathways in different peripheral insulin-sensible tissues (30, 38, 39). Thus, the increase in the number of macrophages, as well as increased proinflammatory adipokines secreted, are hallmarks of the adipose tissue inflammation that accompanies obesity and is associated with the development of insulin resistance and metabolic disease (32, 33, 38).

Adipose Tissue, Free Fatty Acids, and Insulin Resistance

A relevant mechanism linking obesity, insulin resistance, and T2D is the increased release of free fatty acids (FFA) in the circulation by adipose tissue. Under physiological conditions, insulin promotes the increase of glucose uptake, triglyceride synthesis, and repression of lipolysis, a process resulting in the hydrolysis of triglycerides into FFA and glycerol that are released into the circulation (40). However, once the adipose tissue expands, as in cases of obesity, excess lipids and toxic lipid metabolites including FFA, diacylglycerol, and ceramide accumulate in other metabolic tissues, leading to ectopic fat deposition and lipid-induced toxicity (lipotoxicity), and development of insulin resistance in muscle and liver (41, 42). It has been shown that individuals with obesity and T2D have elevated FFA levels in circulation (43, 44), and it is known that circulating FFAs cause insulin resistance in a dose-dependent manner in skeletal muscle and liver (45). The insulin resistance in these peripheral tissues caused by increased levels of FFA will contribute to the loss of glycemic homeostasis.

Mitochondrial Dysfunction and Insulin Resistance, and Adipose Tissue

Mitochondria are highly dynamic intracellular organelles with multiple essential functions and play a critical role in energy metabolism. The key function of mitochondria is cellular respiration, which involves various processes, including activities of mitochondrial electron transport chain complexes and substrate oxidation through the tricarboxylic acid cycle, β-oxidation, ketogenesis, ATP synthesis, and reactive oxygen species (ROS) formation (46–49). An important mechanism linking obesity to diabetes is mitochondrial dysfunction, which leads to impairments in insulin sensitivity in target tissues and compromises pancreatic β-cell function (50). Mitochondrial dysfunction is a broad term that has been used to refer to numerous mitochondrial phenotypes, including decreased respiratory capacity and ATP production, reduced mitochondrial number, accumulated mitochondrial damage due to defects in mitophagy, and altered morphology resulting from changes in mitochondrial fission-fusion dynamic (51). In fact, growing evidence strongly supports the association of reduced mitochondrial function and an increase of reactive oxygen species leading to oxidative stress, particularly in insulin-responsive tissues such as skeletal muscle, white adipose tissue, and the liver (52-54). Particularly in adipose tissue, the mitochondrial number and activity determine the critical threshold at which FFA are released into circulation and exert their lipotoxic effects, promoting insulin resistance in peripheral tissues (46). Clinical and preclinical studies have also shown a reduction in white adipose tissue mitochondria content and activity in obesity and type 2 diabetes (55–57).

Exercise-Induced Adaptations to White Adipose Tissue

Regular physical activity and exercise have important effects on adipose tissue morphology and function, including distinct changes in WAT and BAT. Regarding WAT, exercise can decrease adipocyte size and reduce lipid content in rodents, resulting in decreased adiposity (58, 59). In addition, exercise increases lipolysis and free fatty acid mobilization, which is important to provide metabolic substrate for increased energy demand during exercise, especially during low- to moderate-intensity activities and increased duration. Increased lipolysis is observed during bouts of both endurance and resistance exercise in nonobese individuals and in individuals with obesity (60, 61).

Exercise increases mitochondrial activity and increases the expression of several important metabolic proteins in white adipose tissue, including glucose transporter type 4 (GLUT4) and peroxisome proliferator-activated receptor γ coactivator 1- α (PGC1 α) (58, 62, 63). Exercise training, even performed over a short period (2 wk), improves adipose tissue metabolism, including increases in glucose uptake in subcutaneous WAT and visceral WAT in both healthy and insulin-resistant individuals (64). Moreover, exercise training-induced decreases in adipocyte size and lipid content and increases in GLUT4 and PGC1α expression have been reported in both scWAT and vWAT (59, 65, 66). Fundamentally, several of these metabolic adaptations to adipose tissue can take place independently of significant weight loss showing that adipose tissue can be an important contributor to metabolic health, regardless of alterations in body weight (66). In rodents, exercise training at room temperature induces a "beiging" of scWAT, characterized by increased thermogenic and mitochondrial genes and the presence of adipocytes with multilocular lipid droplets (58, 67, 68), although this is not seen when mice are exercised at thermoneutrality. Most human studies indicate that there is no exercise-induced beiging of scWAT in humans (69-71).

Some beneficial effects of exercise can be mediated by tissue-to-tissue communication, as observed in adipose-muscle tissue cross talk (72). For instance, our laboratory has reported that transplantation of scWAT from exercise-trained donor mice into sedentary recipient mice results in improved glucose homeostasis in the recipient mice (58, 73); and, more recently, transforming growth factor-β2 (TGF-β2) has been identified as the adipokine responsible for the beneficial effects of exercise on glucose metabolism (74), demonstrating that training-induced changes in adipose tissue may have important metabolic effects on overall metabolic health.

Effects of Exercise on Brown Adipose Tissue

BAT is a metabolically active tissue that burns lipids and carbohydrates to generate heat, and it is characterized by a high density of mitochondria, multilocular lipid droplets, and high expression of the thermogenic protein uncoupling protein 1 (UCP1) (24, 75). Several investigations have examined the effects of exercise training on BAT, with conflicting results. For example, some studies have demonstrated that exercise increases BAT activity (76-78), whereas others indicate that exercise decreases mitochondrial activity in BAT (79-81). Recently, a human randomized controlled trial investigated the effects of a 24-wk exercise intervention combining resistance and endurance training in young sedentary adults. Despite a reduction in adiposity and enhanced muscular and cardiorespiratory fitness, exercise has no effect on BAT volume activity in young sedentary adults (82). Further studies need to be performed to clarify the putative effect of exercise training on BAT activity.

Although exercise does not seem to affect BAT volume or the ability of BAT to take up glucose, recent studies have identified an important role for exercise to promote the endocrine function of BAT through the release of batokines. The term batokines refers to BAT-derived molecules, which encompass a variety of signaling molecules including peptides, metabolites, lipids, or microRNAs, and can affect the physiology of a variety of organ systems and cell types (83). A study from our research group has identified the lipokine 12,13-dihydroxy-9Z-octadecenoic acid (12,13-diHOME), which is secreted from BAT in response to exercise in humans and mice and increases skeletal muscle fatty acid uptake and oxidation (72) and cardiac function (84).

Exercise, Obesity, and Adipose Tissue

Although adipose tissue is directly linked to the detrimental effects of obesity on metabolic health, exercise plays a crucial role in managing these negative effects in this tissue. Exercise exerts significant effects on both white and brown adipose tissue that combat the development of obesity and metabolic disease. In WAT, the exercise-induced adaptations include decreasing adiposity and inflammation and increased lipolysis, insulin sensitivity, and enhanced metabolic activity. In BAT, exercise has an important role in promoting its endocrine function through releasing batokines that can mediate some of the positive effects of exercise (Fig. 1).

SKELETAL MUSCLE

Skeletal muscle makes up \sim 40% of the total body mass in mammals and accounts for ~30% of the resting metabolic rate in adult humans (85). In healthy individuals, muscle accounts for around 80% of glucose disposal under insulinstimulated conditions, as it occurs in the postprandial state (86). Skeletal muscle is considered the main tissue responsible for whole body insulin-stimulated glucose disposal and the major site of peripheral insulin resistance (87).

Skeletal Muscle, Obesity, and Inflammation

There are several mechanisms for the development of obesity-induced insulin resistance in skeletal muscle, and inflammation has been proposed to play a role. Growing evidence indicates obesity-induced inflammation occurs in skeletal muscle through proinflammatory pathways activation with increased immune cell infiltration, particularly macrophages and T lymphocytes (88, 89). In addition, obesity is associated with increased muscle inflammatory gene expression and may alter the secretion of different cytokines/myokines (90–92). Similar to visceral fat, muscle macrophages are increased in obesity in the intermyocellular/ intermuscular adipose tissue (IMAT) between the muscle fibers and in perimuscular adipose tissue (PMAT) (89). These IMAT and PMAT macrophages exhibit a proinflammatory, M1-like phenotype, and contribute to higher levels of proinflammatory cytokines, such as TNFα, IL-6, IL-1β, and C-C motif chemokine 2 (CCL2)/monocyte chemoattractant protein (MCP)-1 in skeletal muscle (93). Thus, in obesity, through the secretion of proinflammatory molecules, immune cells may induce myocyte inflammation, adversely regulate myocyte metabolism, and contribute to local and systemic insulin resistance (35, 88, 94).

Skeletal Muscle, Mitochondrial Health, and Insulin Resistance

Mitochondria are particularly important for skeletal muscle function given the high oxidative demands imposed on

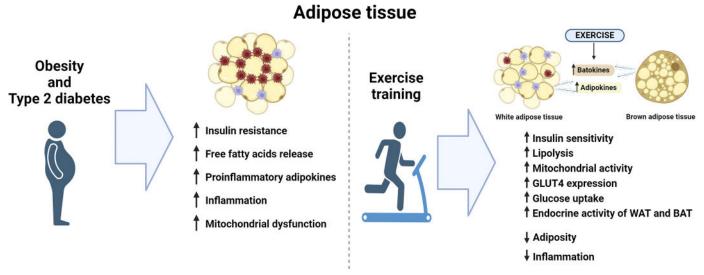


Figure 1. The effects of obesity and type 2 diabetes or exercise on adipose tissue. BAT, brown adipose tissue; GLUT4, glucose transporter type 4; WAT, white adipose tissue. Figure created with Biorender.com.

this tissue by intermittent contraction (47). Mitochondrial health is essential for the proper function of skeletal muscle and metabolic health, and a decline in skeletal muscle mitochondrial content and function is associated with insulin resistance and observed in patients with obesity and type 2 diabetes (46, 95–100), leading to the hypothesis that skeletal muscle mitochondrial dysfunction might be responsible for the development of insulin resistance (101, 102). However, conflicting studies have not observed a reduction in skeletal muscle mitochondria content or function in patients with insulin resistance (103–105). Thus, whether alterations in skeletal muscle mitochondria are a cause or consequence of insulin resistance remain an important topic of discussion (106, 107), the role of mitochondria to contribute to metabolic health is essential. A more comprehensive discussion regarding skeletal muscle mitochondria and insulin resistance is reviewed elsewhere (106, 108, 109).

Exercise-Induced Adaptations to Skeletal Muscle

Regular physical activity and exercise lead to numerous adaptations in skeletal muscle and promote many health benefits, playing a pivotal role in glycemic control and metabolic homeostasis. A well-recognized and important adaptation in skeletal muscle led by exercise is allowing the muscle to become more efficient in generating ATP (11, 110).

In addition, exercise training, especially aerobic exercise training, augments muscle mitochondrial density and function, as well as induces changes in organelle composition (85, 111, 112). For instance, it is well established that 6 wk of aerobic training can increase 50-100% muscle mitochondrial content (113), and training volume and exercise intensity are key determinants of training-induced increases in mitochondrial content and respiration (114). These skeletal muscle exercise-induced adaptations are a hallmark of exercise training and directly contribute to better substrate utilization capacity during exercise, i.e., a decrease in carbohydrate utilization and oxidation and lactate production, and an increase in fat oxidation (110) and insulin sensitivity (115, 116).

The improvement in glycemic homeostasis is a hallmark adaptation of exercise. Exercise training enhances muscle glucose uptake and increases GLUT4 translocation and expression (115, 117). The mechanisms involved in how muscle contraction/exercise increases GLUT4 translocation and expression are complex and are regulated by a combination of several factors, including 5'-AMP-activated protein kinase (AMPK), Ca²⁺/calmodulin-dependent protein kinase II (CaMKII), and RAS-related C3 botulinum toxin substrate 1 (RAC1), among others (117, 118) for GLUT4 translocation; and activation or inhibition of enhancer and repressor transcription factors upon solute carrier family 2 member 4 (SLC2A4) (gene that codifies GLUT4 protein) (117, 119-121). Importantly, the improvements in glucose metabolism and increases in muscle GLUT4 content after exercise training have been showed not only in healthy individuals but also in individuals with T2D (122–124).

Exercise-Released Myokines

An important skeletal muscle adaptation is increasing the release of myokines into circulation, which could mediate some of the beneficial effects of exercise via muscle-organ cross talk with other tissues (125, 126). Myokines are molecules that are produced, expressed, and released by muscle and exert either autocrine, paracrine, or endocrine effects in target tissues (127). Exercise promotes the release of several myokines that mediate or alter the metabolic function of other tissues, including adipose tissue, liver, bone, brain, among others (128, 129). Currently, several myokines are described as involved in exercise adaptation, and some of them are proposed to facilitate the anti-inflammatory effects of exercise and, therefore, critically counteract insulin resistance and the metabolic dysfunction observed in obesity and type 2 diabetes (130). For example, the first to be discovered and one of the most studied myokine is IL-6 (131, 132). Exercise increases the muscle IL-6 expression and secretion in a muscle-contraction proportional manner, particularly when muscle glycogen content is depleted (125, 127). It has been shown that the myokine IL-6 mediates the exerciseassociated anti-inflammatory effects both acutely with each

bout of exercise and as a consequence of training adaptation, including reduction in visceral adipose tissue mass (128, 133). A comprehensive list of exercise-regulated myokines is reviewed elsewhere (128, 134).

Exercise, Obesity, and Skeletal Muscle

Exercise promotes several positive adaptations in skeletal muscle. These adaptations include increased mitochondrial activity and content, enhanced insulin sensitivity and glucose uptake, and reduced inflammation, all of which are impaired in obesity. Moreover, exercise induces the release of myokines, which act as mediators of intertissue communication and contribute to the overall metabolic benefits of exercise (Fig. 2). Thus, exercise not only enhances the function of skeletal muscle but also exerts important systemic effects, playing an essential role in combating the negative effects of obesity and type 2 diabetes on metabolic health, acting as a potent therapeutic tool.

LIVER

The liver is one of the main metabolic organs and its dysregulation plays an important role in the development of insulin resistance and type 2 diabetes. The liver is responsible for the majority source of endogenous glucose production which, under normal postprandial rise in insulin levels, is reduced by activating hepatic glycogen synthesis and suppressing glycogenolysis and gluconeogenesis (4).

Liver, Obesity, and Inflammation

Obesity-induced inflammation may also be observed in the liver. When the liver is insulin resistant, the inhibitory effects of insulin are impaired whereas the stimulatory effect of the hormone on lipogenesis remains intact, contributing to the development of hyperglycemia and hepatic steatosis (32, 36). Similar to adipose tissue, obesity is associated with increased hepatic inflammation and macrophages are the major source of the proinflammatory cytokines. There are two major forms of macrophages in the liver: Kupffer cells (KCs) and recruited hepatic macrophages (RHMs) (35). In obesity, the number of KCs are relatively unchanged, but there is a large increase in RHMs, which are predominantly proinflammatory (34). Although both KCs and RHMs are highly heterogeneous, RHMs express higher levels of M1-like polarized macrophage markers and proinflammatory gene expression, which is exacerbated in obesity (93). Neutrophils are another cell type that accumulates in the liver during the process of obesity and can participate in hepatic inflammation (135). Therefore, obesity is associated with increased recruitment and activation of liver macrophages, increased inflammatory signaling, and local production of inflammatory cytokines and chemokines, particularly the chemokine C-C motif chemokine 2 (CCL2), that can exert paracrine effects generating insulin resistance in hepatocytes (136, 137).

Effects of Exercise on the Liver

Exercise has an important role to improve metabolic health and leads to several adaptations in metabolic tissues, including the liver. Considering the liver has a central role in endogenous glucose production and represents a key site involved in the development of insulin resistance and type 2 diabetes, a significant amount of literature has focused on the effects of exercise upon regulation of glycemic control and insulin sensitivity. An important exercise adaptation to the liver is to enhance the impaired insulin-induced suppressor effect upon hepatic glucose production in individuals with impaired glucose tolerance (138, 139). This is especially important to individuals with type 2 diabetes. Therefore, exercise training promotes an enhanced hepatic insulin sensitivity in individuals with obesity (140-142) and improved hepatic insulin sensitivity and suppression of hepatic glucose production in individuals with type 2 diabetes (143, 144).

As outlined earlier, obesity and type 2 diabetes are associated with increased deposition of intrahepatic lipids that can lead to nonalcoholic fatty liver disease (NAFLD) (145). In

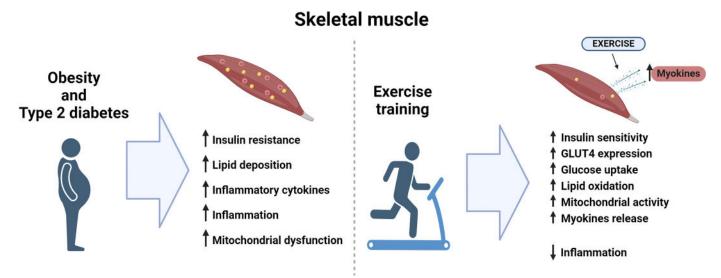


Figure 2. The effects of obesity and type 2 diabetes or exercise on skeletal muscle. GLUT4, glucose transporter type 4. Figure created with Biorender.com.

contrast, exercise training can effectively reduce intrahepatic lipids across multiple populations including individuals with obesity (146, 147), type 2 diabetes (148), and NAFLD (139, 149, 150). Also, the beneficial effects of exercise on reduction in intrahepatic lipids have been observed following different exercise interventions, such as aerobic exercise training, high-intensity intermittent exercise, combined training, among others (151, 152). It is important to mention that the reduction in intrahepatic lipids as an exercise adaptation can be realized in the absence of weight loss, although it is more powerful when significant weight loss is induced (152, 153). Exercise training also leads to increased hepatic fatty acid oxidation, improved mitochondrial function, and increases in other associated mitochondrial outcomes such as beta-hydroxyacyl-CoA dehydrogenase (β-HAD) activity, cytochrome c content, citrate synthase activity, among others in the liver (153).

Exercise-Released Hepatokines

Another relevant role of exercise is promoting the secretion of hepatokines into blood, which can mediate metabolic adaptations to exercise training via liver cross talk with other tissues. Emerging data have identified a significant portion of hepatokines responsive to exercise intervention, but only a few have been functionally linked to the metabolic effects of exercise (10, 154). One of the hepatokines gaining increasing attention due to its potential role in mediating metabolic adaptations to exercise training is fibroblast growth factor 21 (FGF21). Several studies have demonstrated therapeutic benefits of FGF21 for obesity-related metabolic disorders, including the reduction in adiposity and improvement in insulin resistance, NAFLD, among others (155, 156). Clinical and preclinical studies have shown that circulatory levels of FGF21 are increased after acute exercise, whereas decreased after chronic exercise training (≥ 4 wk), due to increased FGF21 sensitivity in adipose tissue, liver, and skeletal muscle (157). It has been suggested in mice that the beneficial effects of exercise-such as the alleviation of obesity-

associated insulin resistance, glucose intolerance, and ectopic lipid accumulation—are abrogated in adipocyte-specific β-klotho (FGF21 receptor) knockout (158), suggesting the important role of FGF21 in mediating the metabolic benefits of exercise (154, 158). A list including other exercise-induced hepatokines related to metabolic diseases is reviewed elsewhere (159, 160).

Obesity, Exercise, and the Liver

Exercise is a potent modulator of hepatic function, leading to key adaptations that enhance metabolic health. The exercise-induced adaptations include enhancement in hepatic insulin sensitivity, effectively suppressing endogenous glucose production and reduction in intrahepatic lipid accumulation. These effects lead to improvements in hepatic lipid oxidation and mitochondrial function and counteract the general effects of obesity on hepatic function. Furthermore, exercise induces the release of hepatokines, which mediate some positive metabolic adaptations and contribute to the systemic benefits of exercise. Future research in this field is needed to elucidate the mechanisms, along with the physiological and clinical implications of exercise-released hepatokines (Fig. 3).

EFFECTS OF EXERCISE IN PATIENTS WITH A GENETIC PREDISPOSITION TO OBESITY

In addition to environmental factors, genetics significantly contribute to the development of obesity, with heritability estimates between 40% and 70% (161). Physical exercise has also been considered a critical preventative tool in people with a genetic predisposition to metabolic disease. For instance, in patients with a genetic predisposition to obesity, exercise interventions can improve cardiorespiratory fitness and muscle strength, alter the biochemical profile (glycemia, lipid profile, and inflammatory markers), and reduce body weight (162, 163). Moreover, important populational studies have shown an inverse association between physical activity and risk for obesity, where increasing

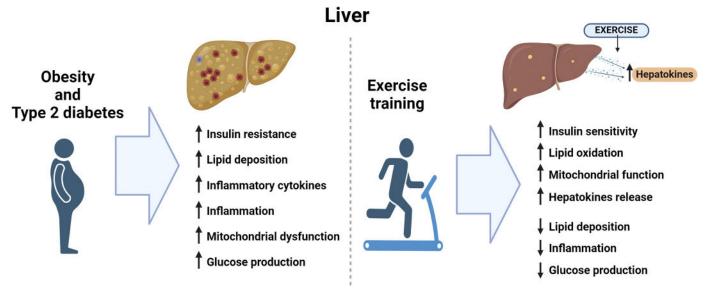


Figure 3. The effects of obesity and type 2 diabetes or exercise on liver. Figure created with Biorender.com.



physical activity can help attenuate the genetic predisposition to obesity (164, 165). A recent study involving over 3,000 participants has found that people with an increased genetic risk of obesity require more exercise (2,300 extra steps per day) to mitigate the risk of obesity (166).

LIMITATIONS

Exercise and metabolic disease encompass broad fields of study, and some of the significant topics had to be summarized or even omitted to be within the scope of this review. For example, the discussion on mitochondrial metabolism and dysfunction was condensed, emphasizing their roles in exercise and insulin resistance. In addition, the genetic predisposition to metabolic disease and the role of exercise were only minimally discussed. Furthermore, only a select number of signaling molecules, such as adipokines, batokines, myokines, hepatokines, and exerkines, and their role in metabolic disease/health were included in this review.

CONCLUSIONS

In summary, several studies have shown how metabolic diseases, particularly obesity and type 2 diabetes, negatively impact metabolic health. Conversely, exercise training emerges as a pivotal preventive and therapeutic strategy with numerous positive effects, exerting profound influences across metabolic tissues. The exercise-induced beneficial adaptations in adipose tissue, skeletal muscle, and liver include enhanced lipolysis, mitochondrial activity, insulin sensitivity, glucose uptake, and reduction of intrahepatic lipids, directly contributing to the improvement of glycemic homeostasis. Furthermore, exercise-induced adaptations can be mediated by the release of molecules from metabolic tissues, termed exerkines, including adipokines from white adipose tissues, batokines from brown adipose tissue, myokines from skeletal muscle, and hepatokines from the liver. These molecules act through endocrine, paracrine, and/or autocrine pathways, facilitating tissue-to-tissue communication. Future research needs to be done to better understand the molecular mechanisms involved in the beneficial exercise-induced adaptations, and the exploration of exerkines presents a promising avenue for understanding and optimizing the therapeutic potential of exercise in metabolic health.

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DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the authors.

AUTHOR CONTRIBUTIONS

J.V.E. and K.I.S. conceived and designed research; J.V.E. prepared figures; J.V.E. and K.I.S. drafted manuscript; J.V.E. and K.I.S. edited and revised manuscript; J.V.E. and K.I.S. approved final version of manuscript.

REFERENCES

- Chew NWS, Ng CH, Tan DJH, Kong G, Lin C, Chin YH, Lim WH, Huang DQ, Quek J, Fu CE, Xiao J, Syn N, Foo R, Khoo CM, Wang JW, Dimitriadis GK, Young DY, Siddiqui MS, Lam CSP, Wang Y, Figtree GA, Chan MY, Cummings DE, Noureddin M, Wong VW, Ma RCW, Mantzoros CS, Sanyal A, Muthiah MD. The global burden of metabolic disease: data from 2000 to 2019. Cell Metab 35: 414-428.e3, 2023. doi:10.1016/j.cmet.2023.02.003.
- International Diabetes Federation. IDF Diabetes Atlas (Online). www.diabetesatlas.org [31 Jan 2024].
- WHO. Obesity and Overweight (Online). https://www.who.int/newsroom/fact-sheets/detail/obesity-and-overweight [2023 Mar 11].
- Lee IM, Shiroma EJ, Lobelo F, Puska P, Blair SN, Katzmarzyk PT; Lancet Physical Activity Series Working Group. Effect of physical inactivity on major non-communicable diseases worldwide: an analysis of burden of disease and life expectancy. Lancet 380: 219-229, 2012. doi:10.1016/S0140-6736(12)61031-9.
- Healy GN, Matthews CE, Dunstan DW, Winkler EA, Owen N. Sedentary time and cardio-metabolic biomarkers in US adults: NHANES 2003-06. Eur Heart J 32: 590-597, 2011. doi:10.1093/ eurhearti/ehq451
- Ozemek C, Lavie CJ, Rognmo Ø. Global physical activity levels need for intervention. Prog Cardiovasc Dis 62: 102-107, 2019. doi:10.1016/j.pcad.2019.02.004.
- Umpierre D, Ribeiro PA, Kramer CK, Leitão CB, Zucatti AT, Azevedo MJ, Gross JL, Ribeiro JP, Schaan BD. Physical activity advice only or structured exercise training and association with HbA1c levels in type 2 diabetes: a systematic review and meta-analysis. JAMA 305: 1790-1799, 2011. doi:10.1001/jama.2011.576.
- Zheng C, Chen XK, Tian XY, Ma AC, Wong SH. Does the gut microbiota contribute to the antiobesity effect of exercise? A systematic review and meta-analysis. Obesity (Silver Spring) 30: 407-423, 2022. doi:10.1002/oby.23345.
- Felix-Soriano E, Stanford KI. Exerkines and redox homeostasis. Redox Biol 63: 102748, 2023. doi:10.1016/j.redox.2023.102748.
- Chow LS, Gerszten RE, Taylor JM, Pedersen BK, van Praag H, Trappe S, Febbraio MA, Galis ZS, Gao Y, Haus JM, Lanza IR, Lavie CJ, Lee CH, Lucia A, Moro C, Pandey A, Robbins JM, Stanford KI, Thackray AE, Villeda S, Watt MJ, Xia A, Zierath JR, Goodpaster BH, Snyder MP. Exerkines in health, resilience and disease. Nat Rev Endocrinol 18: 273-289, 2022. doi:10.1038/s41574-022-00641-2.
- Hawley JA, Hargreaves M, Joyner MJ, Zierath JR. Integrative biology of exercise. Cell 159: 738-749, 2014. doi:10.1016/j.cell.2014.10.029.
- Pescatello LS, MacDonald HV, Lamberti L, Johnson BT. Exercise for hypertension: a prescription update integrating existing recommendations with emerging research. Curr Hypertens Rep 17: 87, 2015. doi:10.1007/s11906-015-0600-y.
- Kanaley JA, Colberg SR, Corcoran MH, Malin SK, Rodriguez NR, Crespo CJ, Kirwan JP, Zierath JR. Exercise/physical activity in individuals with type 2 diabetes: a consensus statement from the American College of Sports Medicine. Med Sci Sports Exerc 54: 353-368, 2022. doi:10.1249/MSS.0000000000002800.
- Garber CE, Blissmer B, Deschenes MR, Franklin BA, Lamonte MJ, Lee IM, Nieman DC, Swain DP; American College of Sports Medicine. American College of Sports Medicine position stand. Quantity and quality of exercise for developing and maintaining cardiorespiratory, musculoskeletal, and neuromotor fitness in apparently healthy adults: guidance for prescribing exercise. Med Sci Sports Exerc 43: 1334-1359, 2011. doi:10.1249/MSS.0b013e318213fefb.
- Cornelissen VA, Fagard RH. Effects of endurance training on blood pressure, blood pressure-regulating mechanisms, and cardiovascular risk factors. Hypertension 46: 667-675, 2005. doi:10.1161/01. HYP.0000184225.05629.51.
- Thorogood A, Mottillo S, Shimony A, Filion KB, Joseph L, Genest J, Pilote L, Poirier P, Schiffrin EL, Eisenberg MJ. Isolated aerobic

- exercise and weight loss: a systematic review and meta-analysis of randomized controlled trials. Am J Med 124: 747-755, 2011. doi:10.1016/j.amjmed.2011.02.037.
- Donnelly JE, Blair SN, Jakicic JM, Manore MM, Rankin JW, Smith BK; American College of Sports Medicine. American College of Sports Medicine Position Stand. Appropriate physical activity intervention strategies for weight loss and prevention of weight regain for adults. Med Sci Sports Exerc 41: 459-471, 2009 [Erratum in Med Sci Sports Exerc 41: 1532, 2009]. doi:10.1249/MSS.0b013e3181949333.
- Andreato LV, Esteves JV, Coimbra DR, Moraes AJP, de Carvalho **T.** The influence of high-intensity interval training on anthropometric variables of adults with overweight or obesity: a systematic review and network meta-analysis. Obes Rev 20: 142-155, 2019. doi:10.1111/ obr.12766
- Savikj M, Zierath JR. Train like an athlete: applying exercise interventions to manage type 2 diabetes. Diabetologia 63: 1491-1499, 2020. doi:10.1007/s00125-020-05166-9.
- 20. Nieuwoudt S, Fealy CE, Foucher JA, Scelsi AR, Malin SK, Pagadala M, Rocco M, Burguera B, Kirwan JP. Functional high-intensity training improves pancreatic β -cell function in adults with type 2 diabetes. Am J Physiol Endocrinol Physiol 313: E314-E320, 2017. doi:10.1152/ aipendo 00407 2016.
- 21. Liu Y, Ye W, Chen Q, Zhang Y, Kuo CH, Korivi M. Resistance exercise intensity is correlated with attenuation of HbA1c and insulin in patients with type 2 diabetes: a systematic review and meta-analysis. Int J Environ Res Public Health 16: 140, 2019. doi:10.3390/ijerph16010140.
- 22. Boulé NG, Haddad E, Kenny GP, Wells GA, Sigal RJ. Effects of exercise on glycemic control and body mass in type 2 diabetes mellitus: a meta-analysis of controlled clinical trials. JAMA 286: 1218-1227, 2001. doi:10.1001/jama.286.10.1218.
- 23. Rosen ED, Spiegelman BM. Adipocytes as regulators of energy balance and glucose homeostasis. Nature 444: 847-853, 2006. doi:10.1038/nature05483.
- 24. Vidal P, Stanford KI. Exercise-induced adaptations to adipose tissue thermogenesis. Front Endocrinol (Lausanne) 11: 270, 2020. doi:10.3389/ fendo.2020.00270.
- 25. Stroh AM, Stanford KI. Exercise-induced regulation of adipose tissue. Curr Opin Genet Dev 81: 102058, 2023. doi:10.1016/j.gde.2023. 102058.
- Golozoubova V, Hohtola E, Matthias A, Jacobsson A, Cannon B, Nedergaard J. Only UCP1 can mediate adaptive nonshivering thermogenesis in the cold. FASEB J 15: 2048-2050, 2001. doi:10.1096/ fi 00-0536fie.
- 27. Foster DO, Frydman ML. Brown adipose tissue: the dominant site of nonshivering thermogenesis in the rat. Experientia Suppl 32: 147-151, 1978. doi:10.1007/978-3-0348-5559-4_16.
- Samuel VT, Shulman Gl. The pathogenesis of insulin resistance: integrating signaling pathways and substrate flux. J Clin Invest 126: 12-22, 2016. doi:10.1172/JCI77812.
- 29. Carbone S, Del Buono MG, Ozemek C, Lavie CJ. Obesity, risk of diabetes and role of physical activity, exercise training and cardiorespiratory fitness. Prog Cardiovasc Dis 62: 327-333, 2019. doi:10.1016/ j.pcad.2019.08.004.
- 30. Kahn SE, Hull RL, Utzschneider KM. Mechanisms linking obesity to insulin resistance and type 2 diabetes. Nature 444: 840-846, 2006. doi:10.1038/nature05482.
- Boden G, She P, Mozzoli M, Cheung P, Gumireddy K, Reddy P, Xiang X, Luo Z, Ruderman N. Free fatty acids produce insulin resistance and activate the proinflammatory nuclear factor-kappaB pathway in rat liver. Diabetes 54: 3458-3465, 2005. doi:10.2337/ diabetes.54.12.3458.
- Saltiel AR, Olefsky JM. Inflammatory mechanisms linking obesity and metabolic disease. J Clin Invest 127: 1-4, 2017. doi:10.1172/ JCI92035
- 33. Lumeng CN, Bodzin JL, Saltiel AR. Obesity induces a phenotypic switch in adipose tissue macrophage polarization. J Clin Invest 117: 175-184, 2007. doi:10.1172/JCI29881.
- 34. Rohm TV, Meier DT, Olefsky JM, Donath MY. Inflammation in obesity, diabetes, and related disorders. Immunity 55: 31-55, 2022. doi:10.1016/j.immuni.2021.12.013.
- 35. Lee YS, Wollam J, Olefsky JM. An integrated view of immunometabolism. Cell 172: 22-40, 2018. doi:10.1016/j.cell.2017.12.025.
- McNelis JC, Olefsky JM. Macrophages, immunity, and metabolic disease. Immunity 41: 36-48, 2014. doi:10.1016/j.immuni.2014.05.010.

- 37. Jia Q, Morgan-Bathke ME, Jensen MD. Adipose tissue macrophage burden, systemic inflammation, and insulin resistance. Am J Physiol Endocrinol Physiol 319: E254-E264, 2020. doi:10.1152/ ajpendo.00109.2020.
- 38. Li M, Chi X, Wang Y, Setrerrahmane S, Xie W, Xu H. Trends in insulin resistance: insights into mechanisms and therapeutic strategy. Signal Transduct Target Ther 7: 216, 2022. doi:10.1038/s41392-022-01073-0.
- 39. Kwon H, Pessin JE. Adipokines mediate inflammation and insulin resistance. Front Endocrinol (Lausanne) 4: 71, 2013. doi:10.3389/ fendo.2013.00071.
- 40. Saponaro C, Gaggini M, Carli F, Gastaldelli A. The subtle balance between lipolysis and lipogenesis: a critical point in metabolic homeostasis. Nutrients 7: 9453-9474, 2015. doi:10.3390/nu7115475.
- Goossens GH. The role of adipose tissue dysfunction in the pathogenesis of obesity-related insulin resistance. Physiol Behav 94: 206-218, 2008. doi:10.1016/j.physbeh.2007.10.010.
- 42. Shulman GI. Ectopic fat in insulin resistance, dyslipidemia, and cardiometabolic disease. N Engl J Med 371: 1131-1141, 2014 [Erratum in N Engl J Med 371: 2241, 2014]. doi:10.1056/NEJMra1011035
- Reaven GM, Hollenbeck C, Jeng CY, Wu MS, Chen YD. Measurement of plasma glucose, free fatty acid, lactate, and insulin for 24 h in patients with NIDDM. Diabetes 37: 1020-1024, 1988. doi:10.2337/diabetes.37.8.1020.
- 44. Boden G. Obesity, insulin resistance and free fatty acids. Curr Opin Endocrinol Diabetes Obes 18: 139-143, 2011. doi:10.1097/MED. 0b013e3283444b09.
- 45. Boden G. Obesity and free fatty acids. Endocrinol Metab Clin North Am 37: 635-646, viii-ix, 2008. doi:10.1016/j.ecl.2008.06.007
- Koliaki C, Roden M. Alterations of mitochondrial function and insulin sensitivity in human obesity and diabetes mellitus. Annu Rev Nutr 36: 337-367, 2016. doi:10.1146/annurev-nutr-071715-050656.
- 47. Patti ME, Corvera S. The role of mitochondria in the pathogenesis of type 2 diabetes. Endocr Rev 31: 364-395, 2010. doi:10.1210/ er.2009-0027.
- 48. Spinelli JB, Haigis MC. The multifaceted contributions of mitochondria to cellular metabolism. Nat Cell Biol 20: 745-754, 2018. doi:10.1038/s41556-018-0124-1.
- 49. Keenan SN, Watt MJ, Montgomery MK. Inter-organelle communication in the pathogenesis of mitochondrial dysfunction and insulin resistance. Curr Diab Rep 20: 20, 2020. doi:10.1007/s11892-020-01300-4.
- 50. Eckel RH, Kahn SE, Ferrannini E, Goldfine AB, Nathan DM, Schwartz MW, Smith RJ, Smith SR. Obesity and type 2 diabetes: what can be unified and what needs to be individualized? J Clin Endocrinol Metab 96: 1654–1663, 2011. doi:10.1210/jc.2011-0585.
- James DE, Stöckli J, Birnbaum MJ. The aetiology and molecular landscape of insulin resistance. Nat Rev Mol Cell Biol 22: 751-771, 2021. doi:10.1038/s41580-021-00390-6.
- 52. Sangwung P, Petersen KF, Shulman Gl, Knowles JW. Mitochondrial dysfunction, insulin resistance, and potential genetic implications. Endocrinology 161: bqaa017, 2020. doi:10.1210/endocr/bqaa017.
- Gonzalez-Franquesa A, Patti ME. Insulin resistance and mitochondrial dysfunction. Adv Exp Med Biol 982: 465–520, 2017. doi:10.1007/ 978-3-319-55330-6_25.
- 54. Fisher-Wellman KH, Neufer PD. Linking mitochondrial bioenergetics to insulin resistance via redox biology. Trends Endocrinol Metab 23: 142-153, 2012. doi:10.1016/j.tem.2011.12.008.
- 55. Chattopadhyay M, Khemka VK, Chatterjee G, Ganguly A, Mukhopadhyay S, Chakrabarti S. Enhanced ROS production and oxidative damage in subcutaneous white adipose tissue mitochondria in obese and type 2 diabetes subjects. Mol Cell Biochem 399: 95-103, 2015. doi:10.1007/s11010-014-2236-7.
- 56. Choo HJ, Kim JH, Kwon OB, Lee CS, Mun JY, Han SS, Yoon YS, Yoon G, Choi KM, Ko YG. Mitochondria are impaired in the adipocytes of type 2 diabetic mice. Diabetologia 49: 784-791, 2006. doi:10.1007/s00125-006-0170-2.
- 57. Wilson-Fritch L, Nicoloro S, Chouinard M, Lazar MA, Chui PC, Leszyk J, Straubhaar J, Czech MP, Corvera S. Mitochondrial remodeling in adipose tissue associated with obesity and treatment with rosiglitazone. J Clin Invest 114: 1281-1289, 2004. doi:10.1172/ JCI21752.
- 58. Stanford KI, Middelbeek RJ, Goodyear LJ. Exercise effects on white adipose tissue: beiging and metabolic adaptations. Diabetes 64:

- 2361-2368, 2015 [Erratum in Diabetes 64: 3334, 2015]. doi:10.2337/
- 59. Gollisch KS, Brandauer J, Jessen N, Toyoda T, Nayer A, Hirshman MF, Goodyear LJ. Effects of exercise training on subcutaneous and visceral adipose tissue in normal- and high-fat diet-fed rats. Am J Physiol Endocrinol Physiol 297: E495-E504, 2009. doi:10.1152/ aipendo 90424 2008
- 60. Richterova B, Stich V, Moro C, Polak J, Klimcakova E, Majercik M, Harant I, Viguerie N, Crampes F, Langin D, Lafontan M, Berlan M. Effect of endurance training on adrenergic control of lipolysis in adipose tissue of obese women. J Clin Endocrinol Metab 89: 1325-1331, 2004. doi:10.1210/jc.2003-031001.
- Ormsbee MJ, Thyfault JP, Johnson EA, Kraus RM, Choi MD, Hickner RC. Fat metabolism and acute resistance exercise in trained men. J Appl Physiol (1985) 102: 1767-1772, 2007. doi:10.1152/ japplphysiol.00704.2006.
- Stallknecht B, Vinten J, Ploug T, Galbo H. Increased activities of mitochondrial enzymes in white adipose tissue in trained rats. Am J Physiol Endocrinol Physiol 261: E410-E414, 1991. doi:10.1152/ajpendo. 1991 261 3 F410
- 63. Lehnig AC, Dewal RS, Baer LA, Kitching KM, Munoz VR, Arts PJ, Sindeldecker DA, May FJ, Lauritzen H, Goodyear LJ, Stanford KI. Exercise training induces depot-specific adaptations to white and brown adipose tissue. iScience 11: 425-439, 2019. doi:10.1016/j. isci.2018.12.033.
- 64. Honkala SM, Motiani P, Kivelä R, Hemanthakumar KA, Tolvanen E, Motiani KK, Eskelinen JJ, Virtanen KA, Kemppainen J, Heiskanen MA, Löyttyniemi E, Nuutila P, Kalliokoski KK, Hannukainen JC. Exercise training improves adipose tissue metabolism and vasculature regardless of baseline glucose tolerance and sex. BMJ Open Diabetes Res Care 8: e000830, 2020. doi:10.1136/bmjdrc-2019-000830
- 65. Sutherland LN, Bomhof MR, Capozzi LC, Basaraba SA, Wright DC. Exercise and adrenaline increase PGC-1\alpha mRNA expression in rat adipose tissue. J Physiol 587: 1607-1617, 2009. doi:10.1113/jphysiol. 2008.165464.
- 66. Stanford KI, Goodyear LJ. Exercise regulation of adipose tissue. Adipocyte 5: 153-162, 2016. doi:10.1080/21623945.2016.1191307.
- 67. Trevellin E, Scorzeto M, Olivieri M, Granzotto M, Valerio A, Tedesco L, Fabris R, Serra R, Quarta M, Reggiani C, Nisoli E, Vettor R. Exercise training induces mitochondrial biogenesis and glucose uptake in subcutaneous adipose tissue through eNOS-dependent mechanisms. Diabetes 63: 2800-2811, 2014. doi:10.2337/db13-1234.
- 68. Stanford KI, Goodyear LJ. Muscle-adipose tissue cross talk. Cold Spring Harb Perspect Med 8: a029801, 2018. doi:10.1101/cshperspect. a029801.
- Vosselman MJ, Hoeks J, Brans B, Pallubinsky H, Nascimento EB, van der Lans AA, Broeders EP, Mottaghy FM, Schrauwen P, van Marken Lichtenbelt WD. Low brown adipose tissue activity in endurance-trained compared with lean sedentary men. Int J Obes (Lond) 39: 1696-1702, 2015. doi:10.1038/ijo.2015.130.
- 70. Tsiloulis T, Carey AL, Bayliss J, Canny B, Meex RCR, Watt MJ. No evidence of white adipocyte browning after endurance exercise training in obese men. Int J Obes (Lond) 42: 721-727, 2018. doi:10.1038/ijo.2017.295.
- Brandao CFC, de Carvalho FG, Souza AO, Junqueira-Franco MVM, Batitucci G, Couto-Lima CA, Fett CA, Papoti M, Freitas EC, Alberici LC, Marchini JS. Physical training, UCP1 expression, mitochondrial density, and coupling in adipose tissue from women with obesity. Scand J Med Sci Sports 29: 1699-1706, 2019. doi:10.1111/
- Stanford KI, Lynes MD, Takahashi H, Baer LA, Arts PJ, May FJ, Lehnig AC, Middelbeek RJW, Richard JJ, So K, Chen EY, Gao F, Narain NR, Distefano G, Shettigar VK, Hirshman MF, Ziolo MT, Kiebish MA, Tseng YH, Coen PM, Goodyear LJ. 12,13-diHOME: an exercise-induced lipokine that increases skeletal muscle fatty acid uptake. Cell Metab 27: 1357, 2018. doi:10.1016/j.cmet.2018.04.023.
- Stanford KI, Middelbeek RJ, Townsend KL, Lee MY, Takahashi H, So K, Hitchcox KM, Markan KR, Hellbach K, Hirshman MF, Tseng YH, Goodyear LJ. A novel role for subcutaneous adipose tissue in exercise-induced improvements in glucose homeostasis. Diabetes 64: 2002-2014, 2015. doi:10.2337/db14-0704.
- 74. Takahashi H, Alves CRR, Stanford KI, Middelbeek RJW, Nigro P, Ryan RE, Xue R, Sakaguchi M, Lynes MD, So K, Mul JD, Lee MY,

- Balan E, Pan H, Dreyfuss JM, Hirshman MF, Azhar M, Hannukainen JC, Nuutila P, Kalliokoski KK, Nielsen S, Pedersen BK, Kahn CR, Tseng YH, Goodyear LJ. TGF- $\beta 2$ is an exerciseinduced adipokine that regulates glucose and fatty acid metabolism. Nat Metab 1: 291-303, 2019. doi:10.1038/s42255-018-0030-7.
- Townsend KL, Tseng YH. Brown fat fuel utilization and thermogenesis. Trends Endocrinol Metab 25: 168-177, 2014. doi:10.1016/j.tem.
- 76. Yoshioka K, Yoshida T, Wakabayashi Y, Nishioka H, Kondo M. Effects of exercise training on brown adipose tissue thermogenesis in ovariectomized obese rats. Endocrinol Jpn 36: 403-408, 1989. doi:10.1507/endocrj1954.36.403.
- Ignacio DL, Fortunato RS, Neto RA, da Silva Silvestre DH, Nigro M, Frankenfeld TG, Werneck-de-Castro JP, Carvalho DP. Blunted response of pituitary type 1 and brown adipose tissue type 2 deiodinases to swimming training in ovariectomized rats. Horm Metab Res 44: 797-803, 2012. doi:10.1055/s-0032-1314875.
- Barbosa MA, Guerra-Sá R, De Castro UGM, de Lima WG, Dos Santos RAS, Campagnole-Santos MJ, Alzamora AC. Physical training improves thermogenesis and insulin pathway, and induces remodeling in white and brown adipose tissues. J Physiol Biochem 74: 441-454, 2018. doi:10.1007/s13105-018-0637-x.
- Wu MV, Bikopoulos G, Hung S, Ceddia RB. Thermogenic capacity is antagonistically regulated in classical brown and white subcutaneous fat depots by high fat diet and endurance training in rats: impact on whole-body energy expenditure. J Biol Chem 289: 34129–34140, 2014. doi:10.1074/jbc.M114.591008.
- Motiani P, Virtanen KA, Motiani KK, Eskelinen JJ, Middelbeek RJ, Goodyear LJ, Savolainen AM, Kemppainen J, Jensen J, Din MU, Saunavaara V, Parkkola R, Löyttyniemi E, Knuuti J, Nuutila P, Kalliokoski KK, Hannukainen JC. Decreased insulin-stimulated brown adipose tissue glucose uptake after short-term exercise training in healthy middle-aged men. Diabetes Obes Metab 19: 1379-1388, 2017. doi:10.1111/dom.12947.
- Dewal RS, Stanford KI. Effects of exercise on brown and beige adipocytes. Biochim Biophys Acta Mol Cell Biol Lipids 1864: 71-78, 2019. doi:10.1016/j.bbalip.2018.04.013.
- 82. Martinez-Tellez B, Sanchez-Delgado G, Acosta FM, Alcantara JMA. Amaro-Gahete FJ. Martinez-Avila WD. Merchan-Ramirez E. Muñoz-Hernandez V, Osuna-Prieto FJ, Jurado-Fasoli L, Xu H, Ortiz-Alvarez L, Arias-Tellez MJ, Mendez-Gutierrez A, Labayen I, Ortega FB, Schonke M, Rensen PCN, Aguilera CM, Llamas-Elvira JM, Gil A, Ruiz JR. No evidence of brown adipose tissue activation after 24 weeks of supervised exercise training in young sedentary adults in the ACTIBATE randomized controlled trial. Nat Commun 13: 5259, 2022. doi:10.1038/s41467-022-32502-x.
- 83. Yang FT, Stanford KI. Batokines: mediators of inter-tissue communication (a mini-review). Curr Obes Rep 11: 1-9, 2022. doi:10.1007/ s13679-021-00465-7
- 84. Pinckard KM, Shettigar VK, Wright KR, Abay E, Baer LA, Vidal P, Dewal RS, Das D, Duarte-Sanmiguel S, Hernandez-Saavedra D, Arts PJ, Lehnig AC, Bussberg V, Narain NR, Kiebish MA, Yi F, Sparks LM, Goodpaster BH, Smith SR, Pratley RE, Lewandowski ED, Raman SV, Wold LE, Gallego-Perez D, Coen PM, Ziolo MT, Stanford KI. A novel endocrine role for the BAT-released lipokine 12,13-diHOME to mediate cardiac function. Circulation 143: 145-159, 2021. doi:10.1161/CIRCULATIONAHA.120.049813.
- 85. Egan B, Zierath JR. Exercise metabolism and the molecular regulation of skeletal muscle adaptation. Cell Metab 17: 162-184, 2013. doi:10.1016/j.cmet.2012.12.012.
- DeFronzo RA, Jacot E, Jequier E, Maeder E, Wahren J, Felber JP. The effect of insulin on the disposal of intravenous glucose. Results from indirect calorimetry and hepatic and femoral venous catheterization. Diabetes 30: 1000-1007, 1981. doi:10.2337/diab.30.12.1000.
- 87. Zierath JR, Krook A, Wallberg-Henriksson H. Insulin action and insulin resistance in human skeletal muscle. Diabetologia 43: 821-835, 2000. doi:10.1007/s001250051457.
- 88. Wu H, Ballantyne CM. Skeletal muscle inflammation and insulin resistance in obesity. J Clin Invest 127: 43-54, 2017. doi:10.1172/ JCI88880.
- 89. Khan IM, Perrard XY, Brunner G, Lui H, Sparks LM, Smith SR, Wang X, Shi ZZ, Lewis DE, Wu H, Ballantyne CM. Intermuscular and perimuscular fat expansion in obesity correlates with skeletal

- muscle T cell and macrophage infiltration and insulin resistance. Int J Obes (Lond) 39: 1607-1618, 2015. doi:10.1038/ijo.2015.104.
- 90. Fink LN, Oberbach A, Costford SR, Chan KL, Sams A, Blüher M, Klip A. Expression of anti-inflammatory macrophage genes within skeletal muscle correlates with insulin sensitivity in human obesity and type 2 diabetes. Diabetologia 56: 1623-1628, 2013. doi:10.1007/ s00125-013-2897-x
- 91. Fink LN, Costford SR, Lee YS, Jensen TE, Bilan PJ, Oberbach A, Bluher M, Olefsky JM, Sams A, Klip A. Pro-inflammatory macrophages increase in skeletal muscle of high fat-fed mice and correlate with metabolic risk markers in humans. Obesity (Silver Spring) 22: 747-757, 2014. doi:10.1002/oby.20615.
- Eckardt K, Görgens SW, Raschke S, Eckel J. Myokines in insulin resistance and type 2 diabetes. Diabetologia 57: 1087-1099, 2014. doi:10.1007/s00125-014-3224-x.
- Lee YS, Olefsky J. Chronic tissue inflammation and metabolic disease. Genes Dev 35: 307-328, 2021. doi:10.1101/gad.346312.120.
- Patsouris D, Cao JJ, Vial G, Bravard A, Lefai E, Durand A, Durand C, Chauvin MA, Laugerette F, Debard C, Michalski MC, Laville M, Vidal H, Rieusset J. Insulin resistance is associated with MCP1-mediated macrophage accumulation in skeletal muscle in mice and humans. PLoS One 9: e110653, 2014. doi:10.1371/journal.pone.0110653.
- 95. Tran L, Langlais PR, Hoffman N, Roust L, Katsanos CS. Mitochondrial ATP synthase β -subunit production rate and ATP synthase specific activity are reduced in skeletal muscle of humans with obesity. Exp Physiol 104: 126-135, 2019. doi:10.1113/EP087278.
- 96. Fabbri E, Chia CW, Spencer RG, Fishbein KW, Reiter DA, Cameron D Zane AC Moore ZA Gonzalez-Freire M Zoli M Studenski SA Kalyani RR, Egan JM, Ferrucci L. Insulin resistance is associated with reduced mitochondrial oxidative capacity measured by 31pmagnetic resonance spectroscopy in participants without diabetes from the baltimore longitudinal study of aging. Diabetes 66: 170-176, 2017. doi:10.2337/db16-0754.
- 97. Mogensen M, Sahlin K, Fernström M, Glintborg D, Vind BF, Beck-Nielsen H, Højlund K. Mitochondrial respiration is decreased in skeletal muscle of patients with type 2 diabetes. Diabetes 56: 1592-1599, 2007. doi:10.2337/db06-0981.
- Chomentowski P, Coen PM, Radiková Z, Goodpaster BH, Toledo FG. Skeletal muscle mitochondria in insulin resistance: differences in intermyofibrillar versus subsarcolemmal subpopulations and relationship to metabolic flexibility. J Clin Endocrinol Metab 96: 494-503, 2011. doi:10.1210/jc.2010-0822.
- 99. Kelley DE, He J, Menshikova EV, Ritov VB. Dysfunction of mitochondria in human skeletal muscle in type 2 diabetes. Diabetes 51: 2944-2950, 2002. doi:10.2337/diabetes.51.10.2944.
- 100. Ritov VB, Menshikova EV, He J, Ferrell RE, Goodpaster BH, Kelley DE. Deficiency of subsarcolemmal mitochondria in obesity and type 2 diabetes. Diabetes 54: 8-14, 2005. doi:10.2337/diabetes.54.1.8.
- 101. Lowell BB, Shulman Gl. Mitochondrial dysfunction and type 2 diabetes. Science 307: 384-387, 2005. doi:10.1126/science.1104343.
- Goodpaster BH. Mitochondrial deficiency is associated with insulin resistance. Diabetes 62: 1032-1035, 2013. doi:10.2337/db12-1612.
- 103. Fisher-Wellman KH, Weber TM, Cathey BL, Brophy PM, Gilliam LA, Kane CL, Maples JM, Gavin TP, Houmard JA, Neufer PD. Mitochondrial respiratory capacity and content are normal in young insulin-resistant obese humans. Diabetes 63: 132-141, 2014. doi:10.2337/
- 104. De Feyter HM, van den Broek NM, Praet SF, Nicolay K, van Loon LJ, Prompers JJ. Early or advanced stage type 2 diabetes is not accompanied by in vivo skeletal muscle mitochondrial dysfunction. Eur J Endocrinol 158: 643-653, 2008. doi:10.1530/EJE-07-0756.
- 105. Boushel R, Gnaiger E, Schjerling P, Skovbro M, Kraunsøe R, Dela F. Patients with type 2 diabetes have normal mitochondrial function in skeletal muscle. Diabetologia 50: 790-796, 2007. doi:10.1007/ s00125-007-0594-3.
- Genders AJ, Holloway GP, Bishop DJ. Are alterations in skeletal muscle mitochondria a cause or consequence of insulin resistance? Int J Mol Sci 21: 6948, 2020. doi:10.3390/ijms21186948.
- Petersen MC, Shulman Gl. Mechanisms of insulin action and insulin resistance. Physiol Rev 98: 2133-2223, 2018. doi:10.1152/physrev. 00063.2017.
- 108. Dela F, Helge JW. Insulin resistance and mitochondrial function in skeletal muscle. Int J Biochem Cell Biol 45: 11-15, 2013. doi:10.1016/j. biocel.2012.09.019.

- 109. Frangos SM, Bishop DJ, Holloway GP. Revisiting the contribution of mitochondrial biology to the pathophysiology of skeletal muscle insulin resistance. Biochem J 478: 3809-3826, 2021. doi:10.1042/
- 110. Hargreaves M, Spriet LL. Skeletal muscle energy metabolism during exercise. Nat Metab 2: 817-828, 2020 [Erratum in Nat Metab 2: 990, 2020], doi:10.1038/s42255-020-0251-4.
- Hood DA, Memme JM, Oliveira AN, Triolo M. Maintenance of skeletal muscle mitochondria in health, exercise, and aging. Annu Rev Physiol 81: 19-41, 2019. doi:10.1146/annurev-physiol-020518-114310.
- Holloszy JO. Biochemical adaptations in muscle. Effects of exercise on mitochondrial oxygen uptake and respiratory enzyme activity in skeletal muscle. J Biol Chem 242: 2278-2282, 1967.
- Hood DA. Invited Review: contractile activity-induced mitochondrial biogenesis in skeletal muscle. J Appl Physiol (1985) 90: 1137-1157, 2001. doi:10.1152/jappl.2001.90.3.1137.
- Granata C, Jamnick NA, Bishop DJ. Training-induced changes in mitochondrial content and respiratory function in human skeletal muscle. Sports Med 48: 1809-1828, 2018. doi:10.1007/s40279-018-0936-v.
- Richter EA, Sylow L, Hargreaves M. Interactions between insulin 115. and exercise. Biochem J 478: 3827-3846, 2021. doi:10.1042/ BCJ20210185.
- Grevendonk L, Connell NJ, McCrum C, Fealy CE, Bilet L, Bruls YMH, Mevenkamp J, Schrauwen-Hinderling VB, Jorgensen JA, Moonen-Kornips E, Schaart G, Havekes B, de Vogel-van den Bosch J, Bragt MCE, Meijer K, Schrauwen P, Hoeks J. Impact of aging and exercise on skeletal muscle mitochondrial capacity, energy metabolism, and physical function. Nat Commun 12: 4773, 2021. doi:10.1038/s41467-021-24956-2.
- Richter EA, Hargreaves M. Exercise, GLUT4, and skeletal muscle glucose uptake. Physiol Rev 93: 993-1017, 2013. doi:10.1152/ physrev.00038.2012
- Sylow L, Kleinert M, Richter EA, Jensen TE. Exercise-stimulated glucose uptake—regulation and implications for glycaemic control. Nat Rev Endocrinol 13: 133-148, 2017. doi:10.1038/nrendo.2016.162.
- Flores-Opazo M, McGee SL, Hargreaves M. Exercise and GLUT4. Exerc Sport Sci Rev 48: 110-118, 2020. doi:10.1249/JES. 0000000000000224.
- Esteves JV, Yonamine CY, Machado UF. SLC2A4 expression and its epigenetic regulation as biomarkers for insulin resistance treatment in diabetes mellitus. Biomark Med 14: 413-416, 2020. doi:10.2217/bmm-2019-0481
- Esteves JV, Enguita FJ, Machado UF. MicroRNAs-mediated regulation of skeletal muscle GLUT4 expression and translocation in insulin resistance. J Diabetes Res 2017: 7267910, 2017. doi:10.1155/2017/ 7267910
- Little JP, Gillen JB, Percival ME, Safdar A, Tarnopolsky MA, Punthakee Z, Jung ME, Gibala MJ. Low-volume high-intensity interval training reduces hyperglycemia and increases muscle mitochondrial capacity in patients with type 2 diabetes. J Appl Physiol (1985) 111: 1554-1560, 2011. doi:10.1152/japplphysiol.00921.2011.
- 123. Hussey SE, McGee SL, Garnham A, Wentworth JM, Jeukendrup AE, Hargreaves M. Exercise training increases adipose tissue GLUT4 expression in patients with type 2 diabetes. Diabetes Obes Metab 13: 959-962, 2011. doi:10.1111/j.1463-1326.2011.01426.x.
- 124. Holten MK, Zacho M, Gaster M, Juel C, Wojtaszewski JF, Dela F. Strength training increases insulin-mediated glucose uptake, GLUT4 content, and insulin signaling in skeletal muscle in patients with type 2 diabetes. Diabetes 53: 294-305, 2004. doi:10.2337/diabetes.53.2.294.
- 125. Laurens C, Bergouignan A, Moro C. Exercise-released myokines in the control of energy metabolism. Front Physiol 11: 91, 2020. doi:10.3389/fphys.2020.00091.
- Das DK, Graham ZA, Cardozo CP. Myokines in skeletal muscle physiology and metabolism: recent advances and future perspectives. Acta Physiol (Oxf) 228: e13367, 2020. doi:10.1111/apha.13367.
- 127. **Pedersen BK**, **Febbraio MA**. Muscles, exercise and obesity: skeletal muscle as a secretory organ. Nat Rev Endocrinol 8: 457-465, 2012. doi:10.1038/nrendo.2012.49.
- Severinsen MCK, Pedersen BK. Muscle-organ crosstalk: the emerging roles of myokines. Endocr Rev 41: 594-609, 2020 [Erratum in Endocr Rev 42: 97-99, 2021]. doi:10.1210/endrev/bnaa016.
- 129. Pedersen BK. Physical activity and muscle-brain crosstalk. Nat Rev Endocrinol 15: 383-392, 2019. doi:10.1038/s41574-019-0174-x.

- 130. Eckel J. Myokines in metabolic homeostasis and diabetes. Diabetologia 62: 1523-1528, 2019. doi:10.1007/s00125-019-4927-9.
- Steensberg A, van Hall G, Osada T, Sacchetti M, Saltin B, Klarlund Pedersen B. Production of interleukin-6 in contracting human skeletal muscles can account for the exercise-induced increase in plasma interleukin-6. J Physiol 529: 237-242, 2000. doi:10.1111/j.1469-7793. 2000.00237.x.
- 132. Pedersen BK, Steensberg A, Fischer C, Keller C, Keller P, Plomgaard P, Febbraio M, Saltin B. Searching for the exercise factor: is IL-6 a candidate? J Muscle Res Cell Motil 24: 113-119, 2003. doi:10.1023/a:1026070911202.
- 133. Wedell-Neergaard AS, Lang Lehrskov L, Christensen RH, Legaard GE, Dorph E, Larsen MK, Launbo N, Fagerlind SR, Seide SK, Nymand S, Ball M, Vinum N, Dahl CN, Henneberg M, Ried-Larsen M, Nybing JD, Christensen R, Rosenmeier JB, Karstoft K, Pedersen BK, Ellingsgaard H, Krogh-Madsen R. Exercise-induced changes in visceral adipose tissue mass are regulated by IL-6 signaling: a randomized controlled trial. Cell Metab 29: 844-855.e3, 2019. doi:10.1016/j.cmet.2018.12.007.
- 134. Hoffmann C, Weigert C. Skeletal muscle as an endocrine organ: the role of myokines in exercise adaptations. Cold Spring Harb Perspect Med 7: a029793, 2017. doi:10.1101/cshperspect.a029793.
- 135. Talukdar S, Oh DY, Bandyopadhyay G, Li D, Xu J, McNelis J, Lu M, Li P, Yan Q, Zhu Y, Ofrecio J, Lin M, Brenner MB, Olefsky JM. Neutrophils mediate insulin resistance in mice fed a high-fat diet through secreted elastase. Nat Med 18: 1407-1412, 2012. doi:10.1038/ nm.2885.
- 136. Johnson AM, Olefsky JM. The origins and drivers of insulin resistance. Cell 152: 673-684, 2013. doi:10.1016/j.cell.2013.01.041.
- 137. Obstfeld AE, Sugaru E, Thearle M, Francisco AM, Gayet C, Ginsberg HN, Ables EV, Ferrante AW Jr. C-C chemokine receptor 2 (CCR2) regulates the hepatic recruitment of myeloid cells that promote obesity-induced hepatic steatosis. Diabetes 59: 916-925, 2010. doi:10.2337/db09-1403.
- Gregory JM, Muldowney JA, Engelhardt BG, Tyree R, Marks-Shulman P, Silver HJ, Donahue EP, Edgerton DS, Winnick JJ. Aerobic exercise training improves hepatic and muscle insulin sensitivity, but reduces splanchnic glucose uptake in obese humans with type 2 diabetes. Nutr Diabetes 9: 25, 2019. doi:10.1038/s41387-019-
- 139. Sargeant JA, Gray LJ, Bodicoat DH, Willis SA, Stensel DJ, Nimmo MA, Aithal GP, King JA. The effect of exercise training on intrahepatic triglyceride and hepatic insulin sensitivity: a systematic review and meta-analysis. Obes Rev 19: 1446-1459, 2018. doi:10.1111/ obr.12719.
- 140. **DeFronzo RA**, **Sherwin RS**, **Kraemer N**. Effect of physical training on insulin action in obesity. Diabetes 36: 1379-1385, 1987. doi:10.2337/ diab.36.12.1379.
- 141. Malin SK, Haus JM, Solomon TP, Blaszczak A, Kashyap SR, Kirwan JP. Insulin sensitivity and metabolic flexibility following exercise training among different obese insulin-resistant phenotypes. Am J Physiol Endocrinol Physiol 305: E1292-E1298, 2013. doi:10.1152/ ajpendo.00441.2013.
- 142. Shojaee-Moradie F, Baynes KC, Pentecost C, Bell JD, Thomas EL, Jackson NC, Stolinski M, Whyte M, Lovell D, Bowes SB, Gibney J, Jones RH, Umpleby AM. Exercise training reduces fatty acid availability and improves the insulin sensitivity of glucose metabolism. Diabetologia 50: 404-413, 2007. doi:10.1007/s00125-006-0498-7.
- 143. Kirwan JP, Solomon TP, Wojta DM, Staten MA, Holloszy JO. Effects of 7 days of exercise training on insulin sensitivity and responsiveness in type 2 diabetes mellitus. Am J Physiol Endocrinol Physiol 297: E151-E156, 2009. doi:10.1152/ajpendo.00210.2009.
- 144. Meex RC, Schrauwen-Hinderling VB, Moonen-Kornips E, Schaart G, Mensink M, Phielix E, van de Weijer T, Sels JP, Schrauwen P, Hesselink MK. Restoration of muscle mitochondrial function and metabolic flexibility in type 2 diabetes by exercise training is paralleled by increased myocellular fat storage and improved insulin sensitivity. Diabetes 59: 572-579, 2010. doi:10.2337/db09-1322.
- David-Silva A, Esteves JV, Morais M, Freitas HS, Zorn TM, Correa-Giannella ML, Machado UF. Dual SGLT1/SGLT2 inhibitor phlorizin ameliorates non-alcoholic fatty liver disease and hepatic glucose production in type 2 diabetic mice. Diabetes Metab Syndr Obes 13: 739-751, 2020. doi:10.2147/DMSO.S242282.

- 146. Johnson NA, Sachinwalla T, Walton DW, Smith K, Armstrong A, Thompson MW, George J. Aerobic exercise training reduces hepatic and visceral lipids in obese individuals without weight loss. Hepatology 50: 1105-1112, 2009. doi:10.1002/hep.23129.
- 147. Keating SE, Hackett DA, Parker HM, O'Connor HT, Gerofi JA, Sainsbury A, Baker MK, Chuter VH, Caterson ID, George J, Johnson NA. Effect of aerobic exercise training dose on liver fat and visceral adiposity. J Hepatol 63: 174-182, 2015. doi:10.1016/j.jhep. 2015.02.022.
- Cassidy S, Thoma C, Hallsworth K, Parikh J, Hollingsworth KG, Taylor R, Jakovljevic DG, Trenell MI. High intensity intermittent exercise improves cardiac structure and function and reduces liver fat in patients with type 2 diabetes: a randomised controlled trial. Diabetologia 59: 56-66, 2016. doi:10.1007/s00125-015-3741-2.
- 149. Bacchi E, Negri C, Targher G, Faccioli N, Lanza M, Zoppini G, Zanolin E, Schena F, Bonora E, Moghetti P. Both resistance training and aerobic training reduce hepatic fat content in type 2 diabetic subjects with nonalcoholic fatty liver disease (the RAED2 Randomized Trial). Hepatology 58: 1287-1295, 2013. doi:10.1002/hep.26393.
- Brouwers B, Schrauwen-Hinderling VB, Jelenik T, Gemmink A, Sparks LM, Havekes B, Bruls Y, Dahlmans D, Roden M, Hesselink MKC, Schrauwen P. Exercise training reduces intrahepatic lipid content in people with and people without nonalcoholic fatty liver. Am J Physiol Endocrinol Physiol 314: E165-E173, 2018. doi:10.1152/ ajpendo.00266.2017.
- Ashcroft SP, Stocks B, Egan B, Zierath JR. Exercise induces tissuespecific adaptations to enhance cardiometabolic health. Cell Metab 36: 278-300, 2024. doi:10.1016/j.cmet.2023.12.008.
- 152. Sargeant JA, Bawden S, Aithal GP, Simpson EJ, Macdonald IA, Turner MC, Cegielski J, Smith K, Dorling JL, Gowland PA, Nimmo MA, King JA. Effects of sprint interval training on ectopic lipids and tissue-specific insulin sensitivity in men with non-alcoholic fatty liver disease. Eur J Appl Physiol 118: 817-828, 2018. doi:10.1007/s00421-018-3818-y.
- Thyfault JP, Rector RS. Exercise combats hepatic steatosis: potential mechanisms and clinical implications. Diabetes 69: 517-524, 2020. doi:10.2337/dbi18-0043.
- 154. Jin L, Diaz-Canestro C, Wang Y, Tse MA, Xu A. Exerkines and cardiometabolic benefits of exercise: from bench to clinic. EMBO Mol Med 16: 432-444, 2024. doi:10.1038/s44321-024-00027-z.
- 155. Geng L, Lam KSL, Xu A. The therapeutic potential of FGF21 in metabolic diseases: from bench to clinic. Nat Rev Endocrinol 16: 654-667, 2020, doi:10.1038/s41574-020-0386-0.
- Jin L, Yang R, Geng L, Xu A. Fibroblast growth factor-based pharmacotherapies for the treatment of obesity-related metabolic complications. Annu Rev Pharmacol Toxicol 63: 359-382, 2023. doi:10.1146/ annurev-pharmtox-032322-093904.
- 157. Porflitt-Rodríguez M, Guzmán-Arriagada V, Sandoval-Valderrama R, Tam CS, Pavicic F, Ehrenfeld P, Martinez-Huenchullán S. Effects of aerobic exercise on fibroblast growth factor 21 in overweight and obesity. A systematic review. Metabolism 129: 155137, 2022. doi:10.1016/j.metabol.2022.155137.
- Geng L, Liao B, Jin L, Huang Z, Triggle CR, Ding H, Zhang J, Huang Y, Lin Z, Xu A. Exercise alleviates obesity-induced metabolic dysfunction via enhancing FGF21 sensitivity in adipose tissues. Cell Rep 26: 2738-2752.e4e2734, 2019. doi:10.1016/j.celrep.2019.02.014.
- 159. Ennequin G, Sirvent P, Whitham M. Role of exercise-induced hepatokines in metabolic disorders. Am J Physiol Endocrinol Physiol 317: E11-E24, 2019. doi:10.1152/ajpendo.00433.2018.
- 160. Weigert C, Hoene M, Plomgaard P. Hepatokines-a novel group of exercise factors. Pflugers Arch 471: 383-396, 2019. doi:10.1007/ s00424-018-2216-y.
- 161. Loos RJF, Yeo GSH. The genetics of obesity: from discovery to biology. Nat Rev Genet 23: 120-133, 2022. doi:10.1038/s41576-021-
- 162. Morales JS, Valenzuela PL, Pareja-Galeano H, Rincón-Castanedo C, Rubin DA, Lucia A. Physical exercise and Prader-Willi syndrome: a systematic review. Clin Endocrinol (Oxf) 90: 649-661, 2019. doi:10.1111/cen.13953.
- Reinehr T, Hebebrand J, Friedel S, Toschke AM, Brumm H, Biebermann H, Hinney A. Lifestyle intervention in obese children with variations in the melanocortin 4 receptor gene. Obesity (Silver Spring) 17: 382-389, 2009. doi:10.1038/oby.2008.422.



- 164. Kilpeläinen TO, Qi L, Brage S, Sharp SJ, Sonestedt E, Demerath E et al. Physical activity attenuates the influence of FTO variants on obesity risk: a meta-analysis of 218,166 adults and 19,268 children. PLoS Med 8: e1001116, 2011. doi:10.1371/journal.pmed.1001116.
- 165. Li S, Zhao JH, Luan J, Ekelund U, Luben RN, Khaw KT, Wareham NJ, Loos RJ. Physical activity attenuates the genetic predisposition
- to obesity in 20,000 men and women from EPIC-Norfolk prospective population study. *PLoS Med* 7: e1000332, 2010. doi:10.1371/journal.pmed.1000332.
- 166. Brittain EL, Han L, Annis J, Master H, Hughes A, Roden DM, Harris PA, Ruderfer DM. Physical activity and incident obesity across the spectrum of genetic risk for obesity. *JAMA Netw Open* 7: e243821, 2024. doi:10.1001/jamanetworkopen.2024.3821.